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(54) Title: ENDODERM, CARDIAC AND NEURAL INDUCING FACTORS

(57) Abstract

Novel proteins have been designated "cerberus" and "frzb-1", respectively. Cerberus is expressed as a secreted peptide during embryogenesis of the *Xenopus* embryo, and is expressed specifically in the head organizer region. This new molecule has endodermal, cardiac, and neural tissue inducing activity, that should prove useful in therapeutic, diagnostic, and clinical applications requiring regeneration, differentiation, or repair of these and other tissues. Frzb-1 is a soluble antagonist of growth factors of the Wnt family that acts by binding to Wnt growth factors in the extracellular space. A third novel protein is termed PAPC which promotes the formation of dorsal mesoderm and somites in the embryo.

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**ENDODERM, CARDIAC AND
NEURAL INDUCING FACTORS**

5 Field of the Invention

The invention generally relates to growth factors, neurotrophic factors, and their inhibitors, and more particularly to several new growth factors with neural, endodermal, and cardiac tissue inducing activity, to complexes and compositions including the factors, and to DNA or RNA coding sequences for the factors. Further, one of the novel growth factors should be useful in tumor suppression gene therapy.

This application claims the benefit of U.S. Provisional Application No. 60/020,150, filed June 20, 1996.

This invention was made with Government support under grant contract number HD-21502, awarded by the National Institutes of Health. The Government has certain rights in this invention.

Background of the Invention

Growth factors are substances, such as polypeptide hormones, which affect the growth of defined populations of animal cells *in vivo* or *in vitro*, but which are not nutrient substances. Proteins involved in the growth and differentiation of tissues may promote or inhibit growth, and promote or inhibit differentiation, and thus the general term "growth factor" includes cytokines, trophic factors, and their inhibitors.

Widespread neuronal cell death accompanies normal development of the central and peripheral nervous systems. Studies of peripheral target tissues during development have shown that neuronal cell death results from the competition among neurons for limiting amounts of survivor factors ("neurotrophic factors"). The earliest identified of these, nerve growth factor ("NGF"), is the most fully characterized and has been shown to be essential for the survival of sympathetic and neural crest-derived sensory neurons during early development of both chick and rat.

One family of neurotropic factors are the Wnts, which have dorsal axis-inducing activity. Most of the Wnt proteins are bound to cell surfaces. (See, e.g., Sokol et al., *Science*, 249, pp. 561-564, 1990.) Dorsal axis-inducing activity in *Xenopus* embryos by one member of this family (*Xwnt-8*) was described by Smith and Harland in 1991, *Cell*, 67, pp. 753-765. The authors described using RNA injections as a strategy for identifying endogenous RNAs involved in dorsal patterning to rescue dorsal development in embryos that were ventralized by UV irradiation.

Another member of the growth and neurotropic factor family was subsequently discovered and described by Harland and Smith, which they termed "noggin." (*Cell*, 70, pp. 829-840 (1992).) Noggin is a good candidate to function as a signaling molecule in Nieuwkoop's center, by virtue of its maternal transcripts, and in Spemann's organizer, through its zygotic organizer-specific expression. Besides noggin, other secreted factors may be involved in the organizer phenomenon.

Another *Xenopus* gene designated "chordin" that begins to be expressed in Spemann's organizer and that can completely rescue axial development in ventralized

embryos was described by Sasai et al., *Cell*, 79, pp. 779-790, 1994. In addition to dorsalizing mesoderm, chordin has the ability to induce neural tissue and its activities are antagonized by Bone Morphogenetic Protein-4 (Sasai et al., *Nature*, 376, pp. 333-336, 1995).

Therefore, the dorsal lip or Spemann's organizer of the *Xenopus* embryo is an ideal tissue for seeking novel growth and neurotrophic factors. New growth and neurotrophic factors are useful agents, particularly those that are secreted due to their ability to be used in physiologically active, soluble forms because these factors, their receptors, and DNA or RNA coding sequences therefore and fragments thereof are useful in a number of therapeutic, clinical, research, diagnostic, and drug design applications.

Summary of the Invention

In one aspect of the present invention, the sequence of the novel peptide that can be in substantially purified form is shown by SEQ ID NO:1. The *Xenopus* derived SEQ ID NO:1 has been designated "cerberus," and this peptide is capable of inducing endodermal, cardiac, and neural tissue development in vertebrates when expressed. The nucleotide sequence which, when expressed results in cerberus, is illustrated by SEQ ID NO:2. Since peptides of the invention induce endodermal, cardiac, and neural tissue differentiation in vertebrates, they should be able to be prepared in physiologically active form for a number of therapeutic, clinical, and diagnostic applications.

Cerberus was isolated during a search for molecules expressed specifically in Spemann's organizer containing a secretory signal sequence. In addition to cerberus, two other novel cDNAs were identified.

The *Xenopus* derived peptide that can be deduced from SEQ ID NO:3 encodes a novel protein we had earlier designated as "frazzled," a secreted protein of 318 amino acids that has dorsalizing activity in *Xenopus* embryos. We now designate the novel protein as "frzb-1." The gene for frzb-1 is expressed in many adult tissues of many animals, three of the cDNAs (*Xenopus*, mouse, and human) have been cloned by us. The accession numbers for the *Xenopus*, mouse, and human frzb-1 cDNA sequences of the gene now designated frzb-1 are U68059, U68058, and U68057, respectively. Frzb-1 has some degree of sequence similarity to the *Drosophila* gene frizzled which has been shown to encode a seven-transmembrane protein that can act both as a signalling and as a receptor protein (Vinson et al., *Nature*, 338, pp. 263-264, 1989; Vinson and Adler, *Nature*, 329, pp. 549-551, 1987). Vertebrate homologues of Frizzled have been isolated and they too were found to be anchored to the cell membrane by seven membrane spanning domains (Wang et al., *J. Biol. Chem.*, 271, pp. 4468-4476, 1996). Frzb-1 differs from the frizzled proteins in that it is an entirely soluble, diffusible secreted protein and therefore —suitable— as a therapeutic agent. The nucleotide sequence derived from *Xenopus* that, when expressed, results in frzb-1 protein is illustrated by SEQ ID NO:4. The frzb-1 protein derived from mouse is shown as SEQ ID NO:7, while the mouse frzb-1 nucleotide sequence is SEQ ID NO:8. The human derived frzb-1 protein is illustrated by SEQ ID NO:9, and the human frzb-1 nucleotide sequence is SEQ ID NO:10.

Frzb-1 is an antagonist of Wnts *in vivo*, and thus is believed to find utility as a tumor suppressor gene, since overexpressed Wnt proteins cause cancer. Frzb-1 may also be a useful vehicle for solubilization

and therapeutic delivery of Wnt proteins complexed with it.

The final cDNA isolated containing a signal sequence results in a peptide designated Paraxial Protocadherin (PAPC). The cDNA for PAPC is a divergent member of the cadherin multigene family. PAPC is most related to protocadherin 43 reported by Sano et al., *The EMBO J.*, 12, pp. 2249-2256, 1993. As shown in SEQ ID NO:5, the PAPC gene encodes a transmembrane protein of 896 amino acids, of which 187 are part of an intracellular domain. PAPC is a cell adhesion molecule, and microinjection of PAPC mRNA constructs into *Xenopus* embryos suggest that PAPC acts as a molecule involved in mesoderm differentiation. A soluble form of the PAPC extracellular domain is able to block muscle and mesoderm formation in *Xenopus* embryos. The nucleotide sequence encoding *Xenopus* PAPC is provided in SEQ ID NO:6.

Cerberus, frzb-1, or PAPC or fragments thereof (which also may be synthesized by *in vitro* methods) may be fused (by recombinant expression or *in vitro* covalent methods) to an immunogenic polypeptide and this, in turn, may be used to immunize an animal in order to raise antibodies against the novel proteins. Antibodies are recoverable from the serum of immunized animals. Alternatively, monoclonal antibodies may be prepared from cells from the immunized animal in conventional fashion. Immobilized antibodies are useful particularly in the diagnosis (*in vitro* or *in vivo*) or purification of cerberus, frzb-1, or PAPC.

Substitutional, deletional, or insertional mutants of the novel polypeptides may be prepared by *in vitro* or recombinant methods and screened for immuno-crossreactivity with cerberus, frzb-1, or PAPC and for cerberus antagonist or agonist activity.

Cerberus or frzb-1 also may be derivatized *in vitro* in order to prepare immobilized and labelled proteins, particularly for purposes of diagnosis of insufficiencies thereof, or for affinity purification of antibodies thereto.

Among applications for the novel proteins are tissue replacement therapy and, because frzb-1 is an antagonist of Wnt signaling, tumor suppression therapies. The cerberus receptor may define a novel signalling pathway. In addition, frzb-1 could permit the isolation of novel members of the Wnt family of growth factors.

Brief Description of the Drawings

Figure 1 illustrates the amino acid sequence (SEQ ID NO:1) of the Fig. 2 cDNA clone for cerberus;

Figure 2 illustrates a cDNA clone (SEQ ID NO:2) for cerberus derived from Xenopus. Sense strand is on top (5' to 3' direction) and the antisense strand on the bottom line (in the opposite direction);

Figures 3 and 4 show the amino acid and nucleotide sequence, respectively, of full-length frzb-1 from Xenopus (SEQ ID NOS:3 and 4);

Figures 5 and 6 show the amino acid and nucleotide sequence, respectively, of full-length PAPC from Xenopus (SEQ ID NOS:5 and 6);

Figures 7 and 8 show the amino acid and nucleotide sequence, respectively, of full-length frzb-1 from mouse (SEQ ID NOS:7 and 8); and

Figures 9 and 10 show the amino acid and nucleotide sequence, respectively, of full-length frzb-1 from human (SEQ ID NOS:9 and 10).

Detailed Description of the Preferred Embodiments

Among the several novel proteins and their nucleotide sequences described herein, is a novel endodermal, cardiac, and neural inducing factor in 5 vertebrates that we have named "cerberus." When referring to cerberus, the present invention also contemplates the use of fragments, derivatives, agonists, or antagonists of cerberus molecules. Because cerberus has no homology to any reported growth factors, 10 it is proposed to be the founding member of a novel family of growth factors with potent biological activities, which may be isolated using SEQ ID NO:2.

The amphibian organizer consists of several cell populations with region-specific inducing 15 activities. On the basis of morphogenetic movements, three very different cell populations can be distinguished in the organizer. First, cells with crawling migration movements involute, fanning out to form the prechordal plate. Second, cells involute 20 through the dorsal lip driven by convergence and extension movements, giving rise to the notochord of the trunk. Third, involution ceases and the continuation of mediolateral intercalation movements leads to posterior extension movements and to the formation of the tail 25 notochord and of the chordoneural hinge. The three cell populations correspond to the head, trunk, and tail organizers, respectively.

The cerberus gene is expressed at the right time and place to participate in cell signalling by 30 Spemann's organizer. Specifically, cerberus is expressed in the head organizing region that consists of crawling-migrating cells. The cerberus expressing region corresponds to the prospective foregut, including the liver and pancreas anlage, and the heart mesoderm.

Cerberus expression is activated by chordin, noggin, and organizer-specific homeobox genes.

Our studies were conducted in early embryos of the frog *Xenopus laevis*. The frog embryo is well suited to experiments, particularly experiments pertaining to generating-and-maintaining regional differences within the embryo for determining roles in tissue differentiation. It is easy to culture embryos with access to the embryos even at very early stages of development (preceding and during the formation of body pattern and differentiation) and the embryos are large. The initial work with noggin and chordin also had been in *Xenopus* embryos, and, as predicted, was highly conserved among vertebrates. Predictions based on work with *Xenopus* as to corresponding human noggin were proven true and the ability to clone the gene for human noggin was readily accomplished. (See the description of *Xenopus* work and cloning information in PCT application, published March 17, 1994, WO 9 405 800, and the subsequent human cloning based thereon in the PCT application, also published March 17, 1994, as WO 9 405 791.)

CLONING

The cloning of cerberus, frzb-1, and PAPC resulted from a comprehensive screen for cDNAs enriched in Spemann's organizer. Subtractive differential screening was performed as follows. In brief, poly A⁺ RNA was isolated from 300 dorsal lip and ventral marginal zone (VMZ) explants at stage 10½. After first strand cDNA synthesis approximately 70-80% of common sequences were removed by subtraction with biotinylated VMZ poly A⁺ RNA prepared from 1500 ventral gastrula halves. For differential screening, duplicate filters (2000 plaques per 15 cm plate, a total of 80,000 clones

screened) of an unamplified oriented dorsal lip library were hybridized with radiolabeled dorsal lip or VMZ cDNA. Putative organizer-specific clones were isolated, grouped by sequence analysis from the 5' end and whole-
5 mount in situ hybridization, and subsequently classified into known and new dorsal-specific genes. Rescreening of the library (100,000 independent phages) with a cerberus probe resulted in the isolation of 45 additional clones, 31 of which had similar size as the
10 longest one of the 11 original clones indicating that they were presumably full-length cDNAs. The longest cDNAs for cerberus, frzb-1, and PAPC were completely sequenced.

To explore the molecular complexity of
15 Spemann's organizer we performed a comprehensive differential screen for dorsal-specific cDNAs. The method was designed to identify abundant cDNAs without bias as to their function. As shown in Table 1, five previously known cDNAs and five new ones were isolated,
20 of which three (expressed as cerberus, frzb-1, and PAPC, respectively) had secretory signal sequences.

TABLE 1

Previously Known Genes	Gene Product	No. of Isolates
Chordin	novel secreted protein	70
Goosecoid	homeobox gene	3
5 Pintallavis/XFKH-1	forkhead/transcription factor	2
Xnot-2	homeobox gene	1
Xlim-1	homeobox gene	1
New Genes		
Cerberus	novel secreted protein	11
10 PAPC	cadherin-like/transmembrane	2
Frzb-1	novel secreted protein	1
Sox-2	sry/transcription factor	1
Fkh-like	forkhead/transcription factor	1

The most abundant dorsal-specific cDNA was
15 chordin (chd), with 70 independent isolates. The second
most abundant cDNA was isolated 11 times and named
cerberus (after a mythological guardian dog with
multiple heads). The cerberus cDNA encodes a putative
secreted polypeptide of 270 amino acids, with an amino
20 terminal hydrophobic signal sequence and a carboxy
terminal cysteine-rich region (Fig. 1). Cerberus is
expressed specifically in the head organizer region of
the Xenopus embryo, including the future foregut.

An abundant mRNA found in the dorsal region of
25 the Xenopus gastrula encodes the novel putative secreted
protein we have designated as cerberus. Cerberus mRNA
has potent inducing activity in Xenopus embryos, leading
to the formation of ectopic heads. Unlike other
organizer-specific factors, cerberus does not dorsalize
30 mesoderm and is instead an inhibitor of trunk-tail
mesoderm. Cerberus is expressed in the anterior-most

domain of the gastrula including the leading edge of the deep layer of the dorsal lip a region that, as shown here, gives rise to foregut and midgut endoderm. Cerberus promotes the formation of cement gland,
5 olfactory placodes, cyclopic eyes, forebrain, and duplicated heart and liver (a foregut derivative). Because the pancreas is also derived from this foregut region, it is likely that cerberus induces pancreas in addition to liver. The expression pattern and inducing
10 activities of cerberus suggest a role for a previously neglected region of the embryo, the prospective foregut endoderm, in the induction of the anterior head region of the embryo.

Turning to Fig. 1, *Xenopus cerberus* encodes a
15 putative secreted protein transiently expressed during embryogenesis and the deduced amino acid sequence of *Xenopus cerberus* is shown. The signal peptide sequence and the nine cysteine residues in the carboxy-terminus are indicated in bold. Potential N-linked glycosylation sites are underlined. In database searches the cerberus protein showed limited similarity only to the mammalian Dan protein, a possible tumor suppressor proposed to be a DNA-binding protein.

Cerberus appears to be a pioneer protein, as
25 its amino acid sequence and the spacing of its 9 cysteine residues were not significantly similar to other proteins in the databases (NCBI-Gen Bank release 93.0). We conclude that the second most abundant dorsal-specific cDNA encodes a novel putative secreted factor, which should be the founding member of a novel family of growth factors active in cell differentiation.
30

Cerberus Demarcates an Anterior Organizer Domain. Cerberus mRNA is expressed at low levels in the unfertilized egg, and zygotic transcripts start accumulating at early gastrula. Expression continues
35

during gastrula and early neurula, rapidly declining during neurulation. Importantly, cerberus expression starts about one hour after that of chd, suggesting that cerberus could act downstream of the chd signal.

5 Whole-mount *in situ* hybridizations reveal that expression starts in the yolk endomesodermal cells located in the deep layer of the organizer. The cerberus domain includes the leading edge of the most anterior organizer cells and extends into the lateral 10 mesoderm. The leading edge gives rise to liver, pancreas, and foregut in its midline, and the more lateral region gives rise to heart mesoderm at later stages of development.

15 Fig. 2 sets out the sequence of a full length Xenopus cDNA for cerberus.

This entirely new molecule has demonstrated physiological properties that should prove useful in therapeutic, diagnostic, and clinical applications that require regeneration, differentiation, or repair of 20 tissues, such wound repair, neuronal regenerational or transplantation, supplementation of heart muscle differentiation, differentiation of pancreas and liver, and other applications in which cell differentiation processes are to be induced.

25 The second, novel, secreted protein we have discovered is called "frzb-1," which was shown to be a secreted protein in Xenopus oocyte microinjection experiments. Thus it provides a natural soluble form of the related extracellular domains of Drosophila and 30 vertebrate frizzled proteins. We propose that the latter proteins could be converted into active soluble forms by introducing a stop codon before the first transmembrane domain. We have noted that the cysteine-rich region of frzb-1 and frizzled contains some overall 35 structural homology with Wnt proteins using the Profile

Search homology program (Gribskov, *Meth. Enzymol.*, 183, pp. 146-159, 1990). This had raised the interesting possibility that frzb-1 could interact directly with Wnt growth factors in the extracellular space. This was because we had found that when microinjected into *Xenopus* embryos, frzb-1 constructs have moderate dorsalizing activity, leading to the formation of embryos with enlarged brain and head, and shortened truck. Somatic muscle differentiation, which requires Xwnt-8, was inhibited. In the case of frzb-1, an attractive hypothesis, suggested by the structural homologies, was that it may act as an inhibitor of Wnt-8, a growth factor that has ventralizing activity in the *Xenopus* embryo (Christian and Moon, *Genes Dev.*, 7, pp. 13-28, 1993). We have shown that frzb-1 can interact with Xwnt-8 and Wnt-1, and it is expected that it could also interact with other members of the Wnt family of growth factors, of which at least 15 members exist in mammals. In addition, a possible interaction with Wnts was suggested by the recent discovery that dishevelled, a gene acting downstream of wingless, has strong genetic interaction with frizzled mutants in *Drosophila* (Krasnow et al., *Development*, 121, pp. 4095-4102, 1995). This possibility has been explored in depth (Leyns et al., *Cell*, 88, pp. 747-756, March 21, 1997), because a soluble antagonist of the Wnt family of proteins is expected to be of great therapeutic value. Examples 1 and 2 illustrate tests that show antagonism of Xwnt-8 by binding to frzb-1.

Vertebrate homologues of Frizzled have been isolated and they too are anchored to the cell membrane by seven membrane spanning domains (Wang et al., *J. Biol. Chem.*, 271, pp. 4468-4476, 1996). Frzb-1 differs from the frizzled proteins in that it is an entirely soluble, diffusible secreted protein and

therefore suitable as a therapeutic agent. The nucleotide sequence that when expressed results in frzb-1 protein is illustrated by SEQ ID NO:4.

SEQ ID NO:4 corresponds to the *Xenopus* homolog, but by using it in BLAST searches (and by cloning mouse frzb-1) we had been able to assemble the sequence of the entire mature human frzb-1 protein, SEQ ID NO:9. Indeed, human frzb-1 is encoded in six expressed sequence tags (ESTs) available in Genebank. The human frzb-1 sequence can be assembled by overlapping in the 5' to 3' direction the ESTs with the following accession numbers in Genebank: H18848, R63748, W38677, W44760, H38379, and N71244. No function had yet been assigned to these EST sequences, but we believe and thus propose here that human frzb-1 will have similar functions in cell differentiation to those described above for *Xenopus* frzb-1. The nucleotide sequence of human frzb-1 is shown in SEQ ID NO:10. The mouse frzb-1 protein and nucleotide sequences are provided by SEQ ID NOS:7 and 8, respectively.

In particular, we believe that frzb-1 will prove useful in gene therapy of human cancer cells. In this rapidly developing field, one approach is to introduce vectors expressing anti-sense sequences to block expression of dominant oncogenes and growth factor receptors. Another approach is to produce episomal vectors that will replicate in human cells in a controlled fashion without transforming the cells. For an example of the latter (an episomal expression vector system for human gene therapy), reference is made to U.S. Patent 5,624,820, issued April 29, 1997, inventor Cooper.

Gene therapy now includes uses of human tumor suppression genes. For example, U.S. Patent 5,491,064, issued February 13, 1996, discloses a tumor suppression

gene localized on chromosome 11 and described as potentially useful for gene therapy in cancers deleted or altered in their expression of that gene. Frzb-1 maps to chromosome 2q31-33 and loss of one copy of the 5 2q31-33 and loss of one copy of the 2q arm has been observed with high incidence in lung carcinomas, colo-rectal carcinomas, and neuroblastomas, which has lead to the proposal that the 2q arm carries a tumor suppressor gene. We expect frzb to be a tumor 10 suppressor gene, and thus to be useful in tumor suppression applications.

A number of applications for cerberus and frzb-1 are suggested from their pharmacological (biological activity) properties.

15 For example, the cerberus and frzb-1 cDNAs should be useful as a diagnostic tool (such as through use of antibodies in assays for proteins in cell lines or use of oligonucleotides as primers in a PCR test to amplify those with sequence similarities to the 20 oligonucleotide primer, and to determine how much of the novel protein is present).

Cerberus, of course, might act upon its target 25 cells via its own receptor. Cerberus, therefore, provides the key to isolate this receptor. Since many receptors mutate to cellular oncogenes, the cerberus receptor should prove useful as a diagnostic probe for certain tumor-types. Thus, when one views cerberus as ligand in complexes, then complexes in accordance with the invention include antibody bound to cerberus, 30 antibody bound to peptides derived from cerberus, cerberus bound to its receptor, or peptides derived from cerberus bound to its receptor or other factors. Mutant forms of cerberus, which are either more potent agonists or antagonists, are believed to be clinically useful.

Such complexes of cerberus and its binding protein partners will find uses in a number of applications.

Practice of this invention includes use of an oligonucleotide construct comprising a sequence coding for cerberus or frzb-1 and for a promoter sequence operatively linked in a mammalian or a viral expression vector. Expression and cloning vectors contain a nucleotide sequence that enables the vector to replicate in one or more selected host cells. Generally, in cloning vectors this sequence is one that enables the vector to replicate independently of the host chromosomes, and includes origins of replication or autonomously replicating sequences. The well-known plasmid pBR322 is suitable for most gram negative bacteria, the 2 μ plasmid origin for yeast and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells.

Expression and cloning vectors should contain a selection gene, also termed a selectable marker. Typically, this is a gene that encodes a protein necessary for the survival or growth of a host cell transformed with the vector. The presence of this gene ensures that any host cell which deletes the vector will not obtain an advantage in growth or reproduction over transformed hosts. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, e.g. ampicillin, neomycin, methotrexate or tetracycline, (b) complement auxotrophic deficiencies.

Examples of suitable selectable markers for mammalian cells are dihydrofolate reductase (DHFR) or thymidine kinase. Such markers enable the identification of cells which were competent to take up the cerberus nucleic acid. The mammalian cell transformants are placed under selection pressure which only the transformants are uniquely adapted to survive by virtue

of having taken up the marker. Selection pressure is imposed by culturing the transformants under conditions in which the concentration of selection agent in the medium is successively changed. Amplification is the
5 process by which genes in greater demand for the production of a protein critical for growth are reiterated in tandem within the chromosomes of successive generations of recombinant cells. Increased quantities of cerberus or frzb-1 can therefore be
10 synthesized from the amplified DNA.

For example, cells transformed with the DHFR selection gene are first identified by culturing all of the transformants in a culture medium which contains methotrexate (Mtx), a competitive antagonist of DHFR.
15 An appropriate host cell in this case is the Chinese hamster ovary (CHO) cell line deficient in DHFR activity, prepared and propagated as described by Urlaub and Chasin, *Proc. Nat. Acad. Sci.*, 77, 4216 (1980). The transformed cells then are exposed to increased levels of Mtx. This leads to the synthesis of multiple copies of the DHFR gene and, concomitantly, multiple copies of other DNA comprising the expression vectors, such as the DNA encoding cerberus or frzb-1. Alternatively, host cells transformed by an expression vector comprising DNA
20 sequences encoding cerberus or frzb-1 and aminoglycoside 3' phosphotransferase (APH) protein can be selected by cell-growth in medium containing an aminoglycosidic antibiotic such as kanamycin or neomycin or G418. Because eukaryotic cells do not normally express an endogenous APH activity, genes encoding APH protein, commonly referred to as neo resistant genes, may be used
25 as dominant selectable markers in a wide range of eukaryotic host cells, by which cells transformed by the vector can readily be identified.

Expression vectors, unlike cloning vectors, should contain a promoter which is recognized by the host organism and is operably linked to the cerberus nucleic acid. Promoters are untranslated sequences located upstream from the start codon of a structural gene (generally within about 100 to 1000 bp) that control the transcription and translation of nucleic acid under their control. They typically fall into two classes, inducible and constitutive. Inducible promoters are promoters that initiate increased levels of transcription from DNA under their control in response to some change in culture conditions, e.g. the presence or absence of a nutrient or a change in temperature. At this time a large number of promoters recognized by a variety of potential host cells are well known. These promoters can be operably linked to cerberus encoding DNA by removing them from their gene of origin by restriction enzyme digestion, followed by insertion 5' to the start codon for cerberus or frzb-1.

Nucleic acid is operably linked when it is placed into a functional relationship with another nucleic acid sequence. For example, DNA for a presequence or secretory leader is operably linked to DNA for a polypeptide if it is expressed as a preprotein which participates in the secretion of the polypeptide; a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the sequence; or a ribosome binding site is operably linked to a coding sequence if it is positioned so as to facilitate translation. Generally, operably linked means that the DNA sequences being linked are contiguous and, in the case of a secretory leader, contiguous and in reading phase. Linking is accomplished by ligation at convenient restriction sites. If such sites do not

exit then synthetic oligonucleotide adapters or linkers are used in accord with conventional practice.

Transcription of the protein-encoding DNA in mammalian host cells is controlled by promoters obtained 5 from the genomes of viruses such as polyoma, cytomegalovirus, adenovirus, retroviruses, hepatitis-B virus, and most preferably Simian Virus 40 (SV40), or from heterologous mammalian promoters, e.g. the actin promoter. Of course, promoters from the host cell or 10 related species also are useful herein.

Cerberus and frzb-1 are clearly useful as a component of culture media for use in culturing cells, such as endodermal, cardiac, and nerve cells, *in vitro*. We believe cerberus and frzb-1 will find uses as agents 15 for enhancing the survival or inducing the growth of liver, pancreas, heart, and nerve cells, such as in tissue replacement therapy.

The final cDNA isolated containing a signal sequence results in a peptide designated Paraxial Protocadherin (PAPC). The cDNA for PAPC is a divergent member of the cadherin multigene family. PAPC is most related to protocadherin 43 reported by Sano et al., *The EMBO J.*, 12, pp. 2249-2256, 1993. As shown in SEQ ID NO:5, the PAPC gene encodes a transmembrane protein of 25 896 amino acids, of which 187 are part of an intracellular domain. PAPC is a cell adhesion molecule, and microinjection of PAPC-mRNA constructs into Xenopus embryos suggest that PAPC acts in mesoderm differentiation. The nucleotide sequence encoding 30 Xenopus PAPC is provided in SEQ ID NO:6.

Therapeutic formulations of the novel proteins may be prepared for storage by mixing the polypeptides having the desired degree of purity with optional physiologically acceptable carriers, excipients or 35 stabilizers, in the form of lyophilized cake or aqueous

solutions. Acceptable carriers, excipients or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; anti-
5 oxidants including ascorbic acid; low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin or immunoglobulins. Other components can include glycine, glutamine, asparagine, arginine, or lysine; monosaccharides,
10 disaccharides, and other carbohydrates including glucose, mannose, or dextrins; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; salt-forming counterions such as sodium; and/or nonionic surfactants such as Tween, Pluronics or PEG.

15 Polyclonal antibodies to the novel proteins generally are raised in animals by multiple subcutaneous (sc) or intraperitoneal (ip) injections of cerberus or frzb-1 and an adjuvant. It may be useful to conjugate these proteins or a fragment containing the target amino
20 acid sequence to a protein which is immunogenic in the species to be immunized, e.g., keyhole limpet hemocyanin, serum albumin, bovine thyroglobulin, or soybean trypsin inhibitor using a bifunctional or derivatizing agent, for example, maleimidobenzoyl
25 sulfosuccinimide ester (conjugation through cysteine residues), N-hydroxysuccinimide (through lysine residues), glutaraldehyde, succinic anhydride, SOCl_2 , or $\text{R}^1\text{N} = \text{C} = \text{NR}$.

30 Animals can be immunized against the immunogenic conjugates or derivatives by combining 1 mg or 1 μg of conjugate (for rabbits or mice, respectively) with 3 volumes of Freund's complete adjuvant and injecting the solution intradermally in multiple sites. One month later the animals are boosted with 1/5 to 1/10
35 the original amount of conjugate in Freund's complete

adjuvant by subcutaneous injection at multiple sites. Seven to 14 days later animals are bled and the serum is assayed for anti-cerberus titer. Animals are boosted until the titer plateaus. Preferably, the animal is
5 boosted with the conjugate of the same cerberus or frzb-1 polypeptide, but conjugated to a different protein and/or through a different cross-linking agent. Conjugates also can be made in recombinant cell culture
10 as protein fusions. Also, aggregating agents such as alum are used to enhance the immune response.

Monoclonal antibodies are prepared by recovering spleen cells from immunized animals and immortalizing the cells in conventional fashion, e.g. by fusion with myeloma cells or by EB virus transformation
15 and screening for clones expressing the desired antibody.

Antibodies are useful in diagnostic assays for cerberus, frzb-1, or PAPC or their antibodies and to identify family members. In one embodiment of a receptor binding assay, an antibody composition which binds to all of a selected plurality of members of the cerberus family is immobilized on an insoluble matrix, the test sample is contacted with the immobilized antibody composition in order to adsorb all cerberus
20 family members, and then the immobilized family members are contacted with a plurality of antibodies specific for each member, each of the antibodies being individually identifiable as specific for a predetermined family member, as by unique labels such as
25 discrete fluorophores or the like. By determining the presence and/or amount of each unique label, the relative proportion and amount of each family member can be determined.
30

The antibodies also are useful for the affinity purification of the novel proteins from
35

recombinant cell culture or natural sources. Antibodies that do not detectably cross-react with other growth factors can be used to purify the proteins free from these other family members.

5

EXAMPLE 1**Frzb-1 Antagonizes Xwnt-8 Non-Cell Autonomously**

To test whether frzb-1 can antagonize secondary axes caused by Xwnt-8 after secretion by injected cells, an experimental design was used. Thus, 10 frzb-1 mRNA was injected into each of the four animal blastomeres of eight-cell embryos, and subsequently, a single injection of Xwnt-8 mRNA was given to a vegetal-ventral blastomere at the 16-32 cell stage. In two independent experiments, we found that injection of 15 frzb-1 alone ($n=13$) caused mild dorsalization with enlargement of the cement gland in all embryos and that injection of Xwnt-8 alone ($n=53$) lead to induction of complete secondary axes in 67% of the embryos. However, injection of frzb-1 into animal caps abolished the 20 formation of complete axes induced by Xwnt-8 ($n=27$), leaving only a residual 14% of embryos with very weak secondary axes. The double-injected embryos retained the enlarged cement gland phenotype caused by injection of frzb-1 mRNA alone. Because both mRNAs encode 25 secreted proteins and were microinjected into different cells, we conclude that the antagonistic effects of frzb-1 and Xwnt-8 took place in the extracellular space after these proteins were secreted.

EXAMPLE 2

Membrane-Anchored Wnt-1 Confers Frzb-1 Binding

To investigate a possible interaction between frzb-1 and Wnts, the first step was to insert an HA epitope tag into a Xenopus frzb-1 construct driven by the CMV (cytomegalovirus) promoter. Frzbl-HA was tested in mRNA microinjection assays in Xenopus embryos and found to be biologically active. Conditioned medium from transiently transfected cells contained up to 10 µg/ml of Frzbl-HA (quantitated on Western blots using an HA-tagged protein standard).

Transient transfection of 293 cells has been instrumental in demonstrating interactions between wingless and frizzled proteins. We therefore took advantage of constructs in which Wnt-1 was fused at the amino terminus of CD8, generating a transmembrane protein containing biologically active Wnt-1 exposed to the extracellular compartment. A Wnt1CD8 cDNA construct (a generous gift of Dr. H. Varmus, NIH) was subcloned into the pCDNA (Invitrogen) vector and transfected into 293 cells. After incubation with Frzbl-HA-conditioned medium (overnight at 37°C), intensely labeled cells were observed by immunofluorescence. As a negative control, a construct containing 120 amino acids of Xenopus chordin, an unrelated secreted protein was used. Transfection of this construct produced background binding of Frzbl-HA to the extracellular matrix, both uniform and punctate. Cotransfection of Wnt1CD8 with pCDNA-LacZ showed that transfected cells stained positively for Frzbl-HA and LacZ. Since Wnt1CD8 contains the entire CD8 molecule, a CD8 cDNA was used as an additional negative control. After transfection with LacZ and full-length CE8, Frzbl-HA failed to bind to the transfected cells. Although most of our experiments

were carried out at 37°C, Frzbl-HA-conditioned medium also stained Wnt1CD8-transfected cells after incubation at 4°C for 2 hours.

Attempts to biochemically quantitate the binding of Frzb-1 to Wnt1CD8-transfected cells were unsuccessful due to high background binding to control cultures, presumably due to binding to the extracellular matrix. Thus, we were unable to estimate a K_D for the affinity of the Frzb-1/Wnt-1 interaction. However, when serial dilutions of conditioned medium containing Frzbl-HA were performed (ranging from 2.5×10^{-7} to 1.25×10^{-10} M), staining of Wnt1CD8-transfected cells was found at all concentrations.

Although we have been unable to provide biochemical evidence for direct binding between Wnts and frzb-1, this cell biological assay indicates that Frzbl-HA can bind, directly or indirectly, to Wnt-1 on the cell membrane in the 10^{-10} M range.

It is to be understood that while the invention has been described above in conjunction with preferred specific embodiments, the description and examples are intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims.

It is Claimed:

1. A substantially pure protein characterized by a physiologically active form and comprising an amino acid sequence encoded by the DNA of SEQ ID NO:2.
2. The protein as in claim 1 having neurotrophic, growth or differentiation factor activity.
3. A composition comprising the protein of claim 1 and a physiologically acceptable carrier with which the peptide is admixed.
4. An oligonucleotide construct comprising a sequence coding for a protein and an expression vector operatively linked therewith, the protein having neurotrophic, growth or differentiation factor activity
5 and being expressible from SEQ ID NO:2.
5. The construct as in claim 4 wherein the expression vector is a mammalian or viral expression vector.
6. A substantially pure protein characterized by a physiologically active form and comprising an amino acid sequence encoded by the DNA of SEQ ID NO:4, SEQ ID NO:8, or SEQ ID NO:10.
7. The protein as in claim 6 having neurotrophic, growth or differentiation factor activity.
8. A composition comprising the protein of claim 6 and a physiologically acceptable carrier with which the protein is admixed.

9. An oligonucleotide construct comprising a sequence coding for a protein and an expression vector operatively linked therewith, the protein being expressible from SEQ ID NO:4, SEQ ID NO:8 or SEQ ID
5 NO:10.

10. The construct as in claim 9 wherein the protein is expressible in soluble form.

11. The construct as in claim 9 wherein the expression vector is a mammalian or viral expression vector.

12. A complex comprising a substantially pure frzb-1 protein complexed with at least one Wnt protein.

13. A substantially pure protein characterized by a physiologically active form and comprising an amino acid sequence encoded by the DNA of SEQ ID NO:6.

14. The protein as in claim 13 having mesoderm differentiation activity.

15. A composition comprising the protein of claim 13 and a physiologically acceptable carrier with which the protein is admixed.

MLLNVLRICI IVCLVNDGAG KHSEGRERTK TYSLNSRGYF	40
RKERGARRSK ILLVNTKGLD EPHIGHGDFG LVAELFDSTR	80
THTNRKEPDM NKVKLFSTVA <u>HGNKSARRKA</u> <u>YNGSRRNIFS</u>	120
RRSFDFKRNT VTEKPGAKMF WNNFLVKMNG <u>APQNTSHGSK</u>	160
AQEIMKEACK TLPFTQNIVH ENCDRMVIQN NLCFGKCISL	200
HVPNQQDRRN TCSHCLPSKF TLNHLT <u>NCT</u> GSKNVVVKVVM	240
MVEECTCEAH KSNFHQTAQF NM D TSTTLHH	270

Figure 1
SUBSTITUTE SHEET (RULE 26)

GAATTCCAG CAAGTCGCTC AGAAACACTG CAGGGTCTAG ATATCATACA ATGTTACTAA	60
CTTAAGGGTC GTTCAGCGAG TCTTGTGAC GTCCCAGATC TATAGTATGT TACAATGATT	
ATGTACTCAG GATCTGTATT ATCGTCTGCC TTGTGAATGA TGGAGCAGGA AAACACTCAG	120
TACATGAGTC CTAGACATAA TAGCAGACGG AACACTTACT ACCTCGTCCT TTTGTGAGTC	
AAGGACGAGA AAGGACAAAA ACATATTCAC TTAACAGCAG AGGTTACTTC AGAAAAGAAA	180
TTCCCTGCTCT TTCCCTGTTT TGTATAAGTG AATTGTGTC TCCAATGAAG TCTTTCTTT	
GAGGAGCAGG TAGGAGCAAG ATTCTGCTGG TGAATACTAA AGGTCTTGAT GAACCCACA	240
CTCCTCGTGC ATCCTCGTTC TAAGACGACC ACTTATGATT TCCAGAACTA CTTGGGGTGT	
TTGGGCATGG TGATTTCGC TTAGTAGCTG AACTATTGA TTCCACCCAGA ACACATACAA	300
AACCCGTACC ACTAAAAGCG AATCATCGAC TTGATAAACT AAGGTGGTCT TGTGTATGTT	
ACAGAAAAGA GCCAGACATG AACAAAGTCA AGCTTTCTC AACAGTTGCC CATGGAAACA	360
TGTCTTTCT CGGTCTGTAC TTGTTTCAGT TCGAAAAGAG TTGTCAACGG GTACCTTTGT	
AAAGTGCAAG AAGAAAAGCT TACAATGGTT CTAGAAGGAA TATTTTCCT CGCCGTTCTT	420
TTTCACGTTTC TTCTTTCGA ATGTTACCAA GATCTCCTT ATAAAAGGA GCGGCAAGAA	
TTGATAAAAG AAATACAGAG GTTACTGAAA AGCCTGGTGC CAAGATGTC TGGAACAATT	480
AACTATTTTC TTTATGTCTC CAATGACTTT TCGGACCAACG GTTCTACAAG ACCTTGTAA	
TTTTGGTTAA AATGAATGGA GCCCCACAGA ATACAAGCCA TGGCAGTAAA GCACAGGAAA	540
AAAACCAATT TTACTTACCT CGGGGTGTCT TATGTCGGT ACCGTCATTT CGTGTCCCTT	
TAATGAAAGA AGCTTGAAA ACCTGTTTT TCACTCAGAA TATTGTACAT GAAAATGTG	600
ATTACTTCT TCGAACGTTT TGGAACAAAA AGTGAGTCTT ATAACATGTA CTTTGACAC	
ACAGGATGGT GATACTGAAAC AATCTGTGCT TTGGTAAATG CATCTCTCTC CATGTTCCAA	660
TGTOCTACCA CTATGTCTTG TTAGACACGA AACCATTTAC GTAGAGAGAG GTACAAGGTT	
ATCAGCAAGA TCGACGAAAT ACTTGTCCC ATTGCTTGCC GTCCAAATT ACCCTGAACC	720
TAGTCGTTCT AGCTGCTTA TGAACAAGGG TAACGAACGG CAGGTTAAA TGGGACTTGG	
ACCTGACGCT GAATTGTACT GGATCTAAGA ATGTAGTAAA GGTTGTCTG ATGGTAGAGG	780
TGGACTGCGA CTTAACATGA CCTAGATTCT TACATCATT CCAACAGTAC TACCATCTCC	
AATGCACGTG TGAAGCTCAT AAGAGCAACT TCCACCAAAC TGCACAGTTT AACATGGATA	840
TTACGTGCAC ACTTCGAGTA TTCTCGTTGA AGGTGGTTG ACGTGTCAAA TTGTACCTAT	
CATCTACTAC CCTGCACCAT TAAAGGACTG CCATACAGTA TGGAAATGCC CTTTGTGTTGG	900
GTAGATGATG GGACGTGGTA ATTTCTGAC GGTATGTCT ACCTTTACGG GAAAACAACC	
AATATTTGTT ACATACTATG CATCTAAAGC ATTATGTGCT CTTCTATTTC ATATAACCAC	960
TTATAAACAA TGTATGATAC GTAGATTCG TAATACAACG GAAGATAAAAG TATATTGGTG	
ATGGAATAAG GATTGTATGA ATTATAATTA ACAAAATGGCA TTTTGTGTAA CATGCAAGAT	1020
TACCTTATTC CTAACATACT TAATATTAAT TGTTACCGT AAAACACATT GTACGTTCTA	

Figure 2A

CTCTGTTCCA TCAGTTGCAA GATAAAAGGC AATATTGTT TGACTTTTT TCTACAAAAT GAGACAAGGT AGTCAACGTT CTATTTCCG TTATAAACAA ACTGAAAAAA AGATGTTTA	1080
GAATACCCAA ATATATGATA AGATAATGGG GTCAAAACTG TTAAGGGTA ATGTAATAAT CTTATGGGTT TATATACTAT TCTATTACCC CAGTTTGAC AATTCCCCAT TACATTATTA	1140
AGGGACTAAG TTTGCCAGG AGCAGTGACC CATAACAACC AATCAGCAGG TATGATTAC TCCCTGATTC AACCGGGTCC TCGTCACTGG GTATTGTTGG TTAGTCGTCC ATACTAAATG	1200
TGGTCACCTG TTTAAAAGCA AACATCTTAT TGGTTGCTAT GGGTTACTGC TTCTGGGCAA ACCAGTGGAC AAATTTCGT TTGTAGAATA ACCAACGATA CCCAATGACG AAGACCCGTT	1260
AATGTGTGCC TCATAGGGGG GTTAGTGTGT TGTGTACTGA ATAAATTGTA TTTATTCAT TTACACACGG AGTATCCCCC CAATCACACA ACACATGACT TATTTAACAT AAATAAAGTA	1320
TGTTACAAAA AAAAAGAA ACAATGTTT TTTTTTTT	

Figure 2B

MSRTRKVDSL LLLAIPGLAL LLLLPNAYCAS CEPVRIPMCK SMPWNMTKMP NHLHHSTQAN 60
AILAIEQFEG LLTTECSQDL LFFLCAMYAP ICTIDFQHEP IKPCKSVCER ARAGCEPILI 120
KYRHTWPESL ACEELPVYDR GVCISPEAIV TVEQGTD SMP DFSMDSNNGN CGSGREHCKC 180
KPMKATQKTY LKNYYNYVIR AKVKEVKVKC HDATAIVEVK EILKSSLVNI PKDTVTLYTN 240
SGCLCPQLVA NEEYIIMGYE DKERTRLLL V EGSLAEKWRD RLAKKVKRWD QKLRRPRKSK 300
DPVAPIPNKN SNSRQARS

Figure 3

GAATTCCCTT TCACACAGGA CTCCTGGCAG AGGTGAATGG TTAGCCCTAT GGATTGGTT	60
CTTAAGGGAA AGTGTGCCT GAGGACCGTC TCCACTTACC AATCGGGATA CCTAAACCAA	
TGTTGATTT GACACATGAT TGATTGCTT CAGATAGGAT TGAAGGACTT GGATTTTAT	120
ACAACTAAAA CTGTGTACTA ACTAACGAAA GTCTATCCTA ACTTCCTGAA CCTAAAAATA	
CTAATTCTGC ACTTTAAAT TATCTGAGTA ATTGTCATT TTGTATTGGA TGGGACTAAA	180
GATTAAGACG TGAAAATTAA ATAGACTCAT TAACAAGTAA AACATAACCT ACCCTGATTT	
GATAAACTTA ACTCCTTGCT TTTGACTTGC CCATAAACTA TAAGGTGGGG TGAGTTGTAG	240
CTATTTGAAT TGAGGAACGA AAACTGAACG GGTATTTGAT ATTCCACCCCC ACTCAACATC	
TTGCTTTAC ATGTGCCAG ATTTCCCTG TATTCCCTGT ATTCCCTCTA AAGTAAGCCT	300
AACGAAAATG TACACGGGTC TAAAAGGGAC ATAAGGGACA TAAGGGAGAT TTCATTGGA	
ACACATACAG GTTGGCAGA ATAACAATGT CTCGAACAAG GAAAGTGGAC TCATTACTGC	360
TGTGTATGTC CAACCCGTCT TATTGTTACA GAGCTGTTG CTTTCACCTG AGTAATGACG	
TACTGCCAT ACCTGGACTG GCGCTCTCT TATTACCCAA TGCTTACTGT GCTTCGTGTG	420
ATGACCGGTAA TGGACCTGAC CGCGAAGAGA ATAATGGGTT ACGAATGACA CGAACGACAC	
AGCCTGTGCG GATCCCCATG TGCAAATCTA TGCCATGGAA CATGACCAAG ATGCCAAC	480
TCGGACACGC CTAGGGGTAC ACGTTAGAT ACGGTACCTT GTACTGGTTC TACGGGTTGG	
ATCTCCACCA CAGCACTCAA GCCAATGCCA TCCTGGCAAT TGAACAGTT GAAGGTTTG	540
TAGAGGTGGT GTCGTGAGTT CGGTTACGGT AGGACCGTTA ACTTGTAAA CTTCCAAACG	
TGACCACTGA ATGTAGCCAG GACCTTTGT TCTTCTGTG TGCCATGTAT GCCCCATTT	600
ACTGGTGACT TACATCGGTC CTGGAAAACA AGAAAGACAC ACGGTACATA CGGGGGTAAA	
GTACCATCGA TTTCCAGCAT GAACCAATTAA AGCCTGCAA GTCCGTGTG GAAAGGGCCA	660
CATGGTAGCT AAAGGTGTA CTTGGTTAAT TCGGAACGTT CAGGCACACG CTTTCCCGGT	
GGGCCGGCTG TGAGCCCATT CTCATAAAGT ACCGGCACAC TTGGCCAGAG AGCCTGGCAT	720
CCCGGGCGAC ACTCGGGTAA GAGTATTCA TGGCCGTGTG AACCGGTCTC TCGGACCGTA	
GTGAAGAGCT GCCCCATAT GACAGAGGAG TCTGCATCTC CCCAGAGGCT ATCGTCACAG	780
CACTTCTCGA CGGGCATATA CTGTCTCCTC AGACGTAGAG GGGTCTCCGA TAGCAGTGT	
TGGAACAAGG AACAGATTCA ATGCCAGACT TCTCCATGGA TTCAAACAAT GGAAATTGCG	840
ACCTTGTCC TTGTCTAAGT TACGGTCTGA AGAGGTACCT AAGTTGTTA CCTTTAACGC	
GAAGCGGCAG GGAGCACTGT AAATGCAAGC CCATGAAGGC AACCCAAAAG ACGTATCTCA	900
CTTCGCOGTC CCTCGTGACA TTTACGTTCG GGTACTTCCG TTGGGTTTC TGCATAGAGT	
AGAATAATTA CAATTATGTA ATCAGAGCAA AAGTGAAGA GGTGAAAGTG AAATGCCACG	960
TCTTATTAAT GTTAATACAT TAGTCTCGTT TTCACTTCT CCACCTTCAC TTTACGGTGC	
ACGCAACAGC AATTGTGGAA GTAAAGGAGA TTCTCAAGTC TTCCCTAGTG AACATTCTA	1020
TGCGTTGTGCG TTAACACCTT CATTTCCTCT AAGAGTTCAAG AAGGGATCAC TTGTAAGGAT	

Figure 4A

AAGACACAGT GACACTGTAC ACCAACTCAG GCTGCTTGTG CCCCCAGCTT GTGCCAATG TTCTGTGTCA CTGTGACATG TGGTTGAGTC CGACGAACAC GGGGGTCGAA CAACGGTTAC	1080
AGGAATAACAT AATTATGGGC TATGAAGACA AAGAGCGTAC CAGGCTTCTA CTAGTGGAAAG TCCTTATGTA TTAATACCCG ATACTTCTGT TTCTCGCATG GTCCGAAGAT GATCACCTTC	1140
GATCCTTGGC CGAAAAATGG AGAGATCGTC TTGCTAAGAA AGTCAAGCGC TGGGATCAAA CTAGGAACCG GCTTTTACC TCTCTAGCAG AACGATTCTT TCAGTCGCG ACCCTAGTTT	1200
AGCTTCGACG TCCCAGGAAA AGCAAAGACC CCGTGGCTCC AATTCCAAC AAAAACAGCA TCGAAGCTGC AGGGTCCTT TCGTTCTGG GGCACCGAGG TTAAGGGTTG TTTTGTCGT	1260
ATTCCAGACA AGCGCGTAGT TAGACTAACG GAAAGGTGA TGAAACTCT ATGGACTTTG TAAGGTCTGT TCGCGCATCA ATCTGATTGC CTTTCCACAT ACCTTGAGA TACCTGAAAC	1320
AAACTAAGAT TTGCATTGTT GGAAGAGCAA AAAAGAAATT GCACTACAGC ACGTTATATT TTTGATTCTA AACGTAACAA CCTTCTCGTT TTTCTTAA CGTGATGTCG TGCAATATAA	1380
CTATTGTTA CTACAAGAAG CTGGTTAGT TGATTGTAGT TCTCCTTCC TTCTTTTT GATAACAAAT GATGTTCTTC GACCAAATCA ACTAACATCA AGAGGAAAGG AAGAAAAAAA	1440
TTATAACTAT ATTTGCACGT GTTCCCAGGC AATTGTTTA TTCAACTTCC AGTGACAGAG AATATTGATA TAAACGTGCA CAAGGGTCCG TTAACAAAAT AAGTTGAAGG TCACTGTCTC	1500
CAGTGAUTGA ATGTCCTCAGC CTAAAGAAGC TCAATTCAATT TCTGATCAAC TAATGGTGAC GTCACTGACT TACAGAGTCG GATTTCTCG AGTTAAGTAA AGACTAGTTG ATTACCACTG	1560
AAGTGTGTTGA TACTTGGGA AAGTGAACTA ATTGCAATGG TAAATCAGAG AAAAGTTGAC TTCACAAACT ATGAACCCCT TTCACATTGAT TAACGTTACC ATTTAGTCTC TTTTCAACTG	1620
CAATGTTGCT TTTCTGTAG ATGAACAAGT GAGAGATCAC ATTTAAATGA TGATCACTTT GTTACAACGA AAAGGACATC TACTGTTCA CTCTCTAGTG TAAATTTACT ACTAGTGAAA	1680
CCATTTAATA CTTTCAGCAG TTTAGTTAG ATGACATGTA GGATGCACCT AAATCTAAAT GGTAAATTAT GAAAGTCGTC AAAATCAATC TACTGTACAT CCTACGTGGA TTTAGATTTA	1740
ATTTTATCAT AAATGAAGAG CTGGTTTAGA CTGTATGGTC ACTGTTGGGA AGGTAAATGC TAAAATAGTA TTTACTTCTC GACCAAATCT GACATACCAAG TGACAACCCCT TCCATTTACG	1800
CTACTTTGTC AATTCTGTT TAAAAATTGC CTAAATAAT ATTAAGTCCT AAATAAAAAA GATGAAACAG TTAAGACAAA ATTTTAACG GATTTATTTA TAATTCAAGGA TTTATTTTT	1860
 AAAAAAAAAAA AAAAA TTTTTTTTTT TTTTT	

Figure 4B
SUBSTITUTE SHEET (RULE 26)

MLLFRAIPM	LLLGLMVLQT	DCEIAQYYID	EEEPPGTVIA	VLSQHSIFNT	TDIPATNFRL	60
MKQFNNSLIG	VRESDGQLSI	MERIDREQIC	RQSLHCNLAL	DVVSFSKGHF	KLLNVKVEVR	120
DINDRSPHFP	SEIMHVEVSE	SSSVGTRIPL	EIAIDEDVGS	NSIQNFQISN	NSHFSIDVLT	180
RADGVKYADL	VLMRELDREI	QPTYIMELLA	MDGGVPSLSG	TAVVNIRVLD	FNDNSPVFER	240
STIAVDLVED	APLGYLLLEL	HATDDDEGVN	GEIVYGFSTL	ASQEVRQLFK	INSRTGSVTL	300
EGQVDFETKQ	TYEFEVQAQD	LGPNPLTATC	KVTVHILDVN	DNTPAITITP	TTTVNAGVAY	360
IPETATKENF	IALISTTDRA	SGSNGQVRCT	LYGHEHFKLQ	QAYEDSYMIV	TTSTLDRENI	420
AAYSLTVVAE	DLGFPSLKTK	KYYTVKVSDE	NDNAPVFSKP	QYEASILENN	APGSYITTVI	480
ARDSDSQNG	KVNYRLVDAK	VMGQSLTTFV	SLDADSGVLR	AVRSLDYEKL	KQDFEIEAA	540
DNGIPQLSTR	VQLNLRIVDQ	NDNCPVITNP	LLNNNGSGEVL	LPISAPQNYL	VFQLKAEDSD	600
EGHNSQLFYT	ILRDPSRLFA	INKESGEVFL	KKQLNSDHSE	DLSIVVAVYD	LGRPSLSTNA	660
TVKFILTDSF	PSNVEVVILO	PSAEEQHQID	MSIIFIHAVLA	GGCALLLLAI	FFVACTCKKK	720
AGEFKQVPEQ	HGTCTNEERLL	STPSPQSVSS	SLSQSESQL	SINTESENCS	VSSNQEQQ	780
TGIKHSISVP	SYHTSGWHL	NCAMSISGHS	HMGHISTKVQ	WAKEIVTSMT	VTLILVENQK	840
RRALSSQCRR	KPVLNTOOMNQ	QGSDMPITIS	ATESTRVQKM	GTAHCNMKRA	IDCLTL	

Figure 5
SUBSTITUTE SHEET (RULE 26)

GAATTCCCAG AGATGAACTC CTTGAGATTG TTTAAATGA CTGCAGGTCT GGAAGGATTC
 CTTAAGGGTC TCTACTTGAG GAACTCTAAC AAAATTTACT GACGTCCAGA CCTTCCTAAC 60

ACATTGCCAC ACTGTTCTA GGCGATGAAAA AACTGCAAGT TTCAACTTG TTTTGGTGC
 TGTAACGGTG TGACAAAGAT CCGTACTTT TTGACGTTCA AAGTTGAAAC AAAAACACG 120

-AACTTGATT CTTCAAGATG CTGCTTCTCT TCAGAGCCAT TCCAATGCTG CTGTTGGGAC
 TTGAAACTAA GAAGTTCTAC GACGAAGAGA AGTCTCGGTAGGTTACGAC GACAACCCCTG 180

TGATGGTTTT ACAAACAGAC TGTGAAATTG CCCAGTACTA CATAGATGAA GAAGAACCCCC
 ACTACCAAAA TGTTGTCTG ACACCTTAAC GGGTCATGAT GTATCTACTT CTTCTTGGGG 240

CTGGCACTGT AATTGCAGTG TTGTCACAAC ACTCCATATT TAACACTACA GATATACCTG
 GACCGTGACA TTAACGTCAC AACAGTGTG TGAGGTATAA ATTGTGATGT CTATATGGAC 300

CAACCAATTG CCGTCTAATG AAGCAATTG ATAATTCCCT TATCGGAGTC CGTGAGAGTG
 GTTGGTTAAA GGCAGATTAC TTCGTTAAAT TATTAAGGGA ATAGCCTCAG GCACTCTCAC 360

ATGGGCAGCT GAGCATCATG GAGAGGATTG ACCGGGAGCA AATCTGCAGG CAGTCCTTC
 TACCCGTGCA CTCGTAGTAC CTCTCCTAAC TGGCCCTCGT TTAGACGTCC GTCAGGGAAAG 420

ACTGCAACCT GGCTTGGAT GTGGTCAGCT TTTCAAAGG ACACCTCAAG CTTCTGAACG
 TGACGTTGGA CCGAACCTA CACCAGTCGA AAAGGTTCC TGTGAAGTTC GAAGACTTGC 480

TGAAAGTGGG AGTGAGAGAC ATTAATGACC ATAGCCTCA CTTTCCCAGT GAAATAATGC
 ACTTTCACCT CCACTCTCTG TAATTACTGG TATCGGGAGT GAAAGGGTCA CTTTATTACG 540

ATGTGGAGGT GTCTGAAAGT TCCTCTGTGG GCACCAAGGAT TCCTTAGAA ATTGCAATAG
 TACACCTCCA CAGACTTCA AGGAGACACC CGTGGTCCTA AGGAAATCTT TAACGTTATC 600

ATGAAGATGT TGGGTCCAAC TCCATCCAGA ACTTTAGAT CTCAAATAAT AGCCACTTCA
 TACTTCTACA ACCCAGGTTG AGGTAGGTCT TGAAAGTCTA GAGTTTATTA TCGGTGAAGT 660

GCATTGATGT GCTAACCCAGA GCAGATGGGG TGAAATATGC AGATTAGTC TTAATGAGAG
 CGTAACTACA CGATTGGTCT CGTCTACCCC ACTTTATACG TCTAAATCAG AATTACTCTC 720

AACTGGACAG GGAAATCCAG CCAACATACA TAATGGAGCT ACTAGCAATG GATGGGGGTG
 TTGACCTGTC CCTTTAGGTC GGTTGTATGT ATTACCTCGA TGATCGTTAC CTACCCOCAC 780

TACCATCACT ATCTGGTACT GCAGTGGTTA ACATCCGAGT CCTGGACTTT AATGATAACA
 ATGGTAGTGA TAGACCATGA CGTCACCAAT TGTAGGCTCA GGACCTGAAA TTACTATTGT 840

GCCCAGTGT TGAGAGAAGC ACCATTGCTG TGGACCTAGT AGAGGATGCT CCTCTGGGAT
 CGGGTCACAA ACTCTCTTCG TGGTAACGAC ACCTGGATCA TCTCCTACGA GGAGACCCCTA 900

ACCTTTGTT GGAGTTACAT GCTACTGACG ATGATGAAGG AGTGAATGGA GAAATTGTTT
 TGGAAAACAA CCTCAATGTA CGATGACTGC TACTACTTCC TCACTTACCT CTTAACAAA 960

ATGGATTCAAG CACTTTGGCA TCTCAAGAGG TACGTCAAGCT ATTTAAAATT AACTCCAGAA
 TACCTAAGTC GTGAAACCGT AGAGTTCTCC ATGCAGTCGA TAAATTTAA TTGAGGTCTT 1020

CTGGCAGTGT TACTCTTGAA GGCCAAGTTG ATTTGAGAC CAAGCAGACT TACGAATTTG GACCGTCACA ATGAGAACCT CCGGTTCAAC TAAAACCTCG GTTCGTCTGA ATGCTTAAAC	1080
AGGTACAAGC CCAAGATTTG GGCCCCAACCC CACTGACTGC TACTGTAAA GTAACGTTC TCCATGTTCG GGTTCTAAAC CGGGGGTTGG GTGACTGACG ATGAACATT CATTGACAAG	1140
ATATACTTGA TGTAATGAT AATACCCCAG CCATCACTAT TACCCCTCTG ACTACTGTAA TATATGAACT ACATTTACTA TTATGGGGTC GGTAGTGATA ATGGGGAGAC TGATGACATT	1200
ATGCAGGAGT TGCCTATATT CCAGAAACAG CCACAAAGGA GAACTTTATA GCTCTGATCA TACGTCTCA ACGGATATAA GGTCTTGTG CCGTGGTCCCT CTTGAAATAT CGAGACTAGT	1260
GCACTACTGA CAGAGCCTCT GGATCTAATG GACAAGTTCG CTGTACTCTT TATGGACATG CGTGATGACT GTCTCGGAGA CCTAGATTAC CTGTTCAAGC GACATGAGAA ATACCTGTAC	1320
AGCACTTTAA ACTACAGCAA GCTTATGAGG ACAGTTACAT GATAGTTACC ACCTCTACTT TCGTGAAATT TGATGTCGTT CGAATACTCC TGTCAATGTA CTATCAATGG TGGAGATGAA	1380
TAGACAGGGA AAACATAGCA GCGTACTCTT TGACAGTAGT TGCAGAAGAC CTTGGCTTCC ATCTGTCCTT TTTGTATCGT CGCATGAGAA ACTGTCATCA ACGTCTTCTG GAACCGAAGG	1440
CCTCATTGAA GACCAAAAAG TACTACACAG TCAAGGTTAG TGATGAGAAT GACAATGCAC GGAGTAACCTT CTGGTTTTTC ATGATGTCG AGTTCCAATC ACTACTCTTA CTGTTACGTG	1500
CTGTATTTTC TAAACCCCAG TATGAAGCTT CTATTCTGGA AAATAATGCT CCAGGCTCTT GACATAAAAG ATTTGGGGTC ATACTTCGAA GATAAGACCT TTTATTACGA GGTCCGAGAA	1560
ATATAACTAC AGTGATAGCC AGAGACTCTG ATAGTGATCA AAATGGAAA GTAAATTACA TATATTGATG TCACTATCGG TCTCTGAGAC TATCACTAGT TTTACCGTTT CATTAAATGT	1620
GACTGTGGA TGCAAAAGTG ATGGGCCAGT CACTAACAC ATTTGTTCT CTTGATGCGG CTGAACACCT ACgtttcac TACCCGGTCA GTGATTGTTG TAAACAAAGA GAACTACGCC	1680
ACTCTGGAGT ATTGAGAGCT GTTAGGTCTT TAGACTATGA AAAACTAAA CAACTGGATT TGAGACCTCA TAATCTCGA CAATCCAGAA ATCTGATACT TTTGAATT GTTGACCTAA	1740
TTGAAATTGA AGCTGCAGAC AATGGGATCC CTCAACTCTC CACTCGCGTT CAACTAAATC AACTTTAATC TCGACGTCTG TTACCCCTAGG GAGTTGAGAG GTGAGCGAA GTTGATTTAG	1800
TCAGAATAGT TGATCAAAAT GATAATTGOC CTGTGATAAC TAATCCTCTT CTTAATAATG AGTCTTATCA ACTAGTTTA CTATTAACGG GACACTATTG ATTAGGAGAA GAATTATTAC	1860
GCTGGGTGA AGTTCTGCTT CCCATCAGCG CTCCTCAAAA CTATTTAGTT TTCCAGCTCA CGAGCCCACT TCAAGACGAA GGGTAGTCGC GAGGAGTTT GATAAAATCAA AAGGTCGAGT	1920
AAGCCGAGGA TTCAGATGAA GGGCACAACT CCCAGCTGTT CTATACCATA CTGAGAGATC TTCGGCTCCT AAGTCTACTT CCCGTGTTGA GGGTCGACAA GATATGGTAT GACTCTCTAG	1980
CAAGCAGATT GTTTGCCATT AACAAAGAAA GTGGTGAAGT GTTCCTGAAA AAACAATTAA GTTCGTCTAA CAAACGGTAA TTGTTTCTT CACCACTTC CAAGGACTTT TTTGTTAATT	2040
ACTCTGACCA TTCAGAGGAC TTGAGCATAG TAGTTGCAGT GTATGACTTG GGAAGACCTT TGAGACTGGT AAGTCTCCTG AACTCGTATC ATCAACGTCA CATACTGAAC CCTTCTGGAA	2100
CATTATCCAC CAATGCTACA GTTAAATTCA TCCTCACCGA CTCTTTCTT TCTAACGTG GTAATAGGTG GTTACGATGT CAATTAAAGT AGGAGTGGCT GAGAAAAGGA AGATTGCAAC	2160

AAGTCGTTAT TTTGCAACCA TCTGCAGAAG AGCAGCACCA GATCGATATG TCCATTATAT TTCAGCAATA AAACGTTGGT AGACGTCTTC TCGTCGTGGT CTAGCTATAC AGGTAATATA	2220
TCATTGCAGT GCTGGCTGGT GGTTGTGCTT TGCTACTTTT GGCCATCTTT TTTGTGGCCT AGTAACGTCA CGACCGACCA CCAACACGAA ACGATGAAAA CCGGTAGAAA AACACCCGGA	2280
GTACTTGTAA AAAGAAAGCT GGTGAATTAA AGCAGGTACC TGAACAACAC GGAACATGCA CATGAACATT TTTCTTCGA CCACTTAAAT TCGTCCATGG ACTTGTGTG CCTTGTACGT	2340
ATGAAGAACG CCTGTTAACG ACCCCATCTC CCCAGTCGGT CTCTCTTCT TTGTCTCAGT TACTTCTTGC GGACAATTGAG GGGTCAGCCA GAGAAGAAGA AACAGAGTCA	2400
CTGAGTCATG CCAACTCTCC ATCAAACTG AATCTGAGAA TTGCAGCGTG TCCTCTAAC GACTCAGTAC GGTTGAGAGG TAGTTATGAC TTAGACTCTT AACGTCGCAC AGGAGATTGG	2460
AAGAGCAGCA TCAGCAAACA GGCATAAAGC ACTCCATCTC TGTACCATCT TATCACACAT TTCTCGTCGT AGTCGTTGT CGTATTTCG TGAGGTAGAG ACATGGTAGA ATAGTGTGTA	2520
CTGGTTGGCA CCTGGACAAT TGTGCAATGA GCATAAGTGG ACATTCTCAC ATGGGGCACA GACCAACCCTG GGACCTGTTA ACACGTTACT CGTATTCAACC TGTAAGAGTG TACCCGTGT	2580
TTAGTACAAA GGTACAGTGG GCAAAGGAGA TAGTGAATTC AATGACAGTG ACTCTGATAC AATCATGTTT CCATGTCACC CGTTCCCTCT ATCACTGAAG TTACTGTCAC TGAGACTATG	2640
TAGTGGAGAA TCAGAAAAGA AGAGCATTGA GCAGCCAATG CAGGCACAAAG CCAGTGCTCA ATCACCTCTT AGTCTTTCT TCTCGTAACG CGTCGGTTAC GTCCGTGTTG GGTACACGAGT	2700
ATACACAGAT GAATCAGCAG GGTTCCGACA TGCCGATAAC TATTTCAGCC ACCGAATCAA TATGTGTCTA CTTAGTCGTC CCAAGGCTGT ACGGCTATTG ATAAAGTCGG TGGCTTAGTT	2760
CAAGGGTCCA GAAAATGGGA ACTGCACATT GCAATATGAA AAGGGCTATA GACTGTCTTA GTTCCCAGGT CTTTACCCCT TGACGTGTAA CGTTATACTT TTCCCGATAT CTGACAGAAAT	2820
CTCTGTAGCT CCTGTATATT ACAATACCTA CCATGCAAGA ATGCCTAACCC TGCACATACC GAGACATCGA GGACATATAA TGTTATGGAT GGTACGTTCT TACGGATTGG ACGTGTATGG	2880
GAACCATAACC CTTAGAGACC CTTATTACCA TATCAATAAT CCTGTTGCTA ATCGGATGCA CTTGGTATGG GAATCTCTGG GAATAATGGT ATAGTTATTA GGACAACGAT TAGCCTACGT	2940
GGCGGAATAT GAAAGAGATT TAGTCAACAG AAGTGCAACG TTATCTCCGC AGAGATCGTC CCGCCTTATAA CTTTCTCTAA ATCAGTTGTC TTCACGTTGC AATAGAGGCG TCTCTAGCAG	3000
TAGCAGATAAC CAAGAATTCA ATTACAGTCC GCAGATATCA AGACAGCTTC ATCCTTCAGA ATCGTCTATG GTTCTTAAGT TAATGTCAGG CGTCTATAGT TCTGTCGAAG TAGGAAGTCT	3060
AATTGCTACA ACCTTTAAT CATTAGGCAT GCAAGTGAGA ATGCACAAAG GCAAGTGCTT TTAACGATGT TGGAAAATTA GTAATCGTA CGTTCACTCT TACGTGTTTC CGTTCACGAA	3120
TAGCATGAAA GCTAAATATA TGGAGTCTCC CCTTTCCCTC TGATGGATGG GGGGAGACAC ATCGTACTTT CGATTATAT ACCTCAGAGG GGAAAGGGAG ACTACCTACC CCCCTCTGTG	3180
AGGACAGTGC ATAAATATAC AGCTGTTTC TATTGCTATT TCACTTGGGA ATTTTTGTT TCCTGTCAAG TATTATATG TCGACGAAAG ATAAACGTAAG AGTGAACCCCT TAAAAAAACAA	3240
TTTTTTACAT ATTTATTTT CCTGAATTGA ATGTGACATT GTCCTGTCAC CTAACTAGCA AAAAAATGTA TAAATAAAA GGACTTAAC TACACTGTAA CAGGACAGTG GATTGATCGT	3300

Figure 6C
SUBSTITUTE SHEET (RULE 26)

ATTAATCCA CAGACCTACA GTCAAAATATT TGAGGGCCCC TGAAACAGCA CATCAGTCAG TAATTTAGGT GTCTGGATGT CAGTTATAA ACTCCCGGGG ACTTTGTCGT GTAGTCAGTC	3360
GACCTAAAGT GGCCCTTTTA CTTTAGCAG CTCCCTGGTC TGCCCTCTGT GTTAATCAGC CTGGATTCA CGGGAAAAAT GAAAATCGTC GAGGACCCAG ACGGGAGACA CAATTAGTCG	3420
CCCTGGTCAA GTCTGAGTA GGATCATGGC GTTTTATAT GCATCTCACC TACTTGGAC GGGACCAAGTT CAGGACTCAT CCTAGTACCG CAAAAATAA CGTAGAGTGG ATGAAACCTG	3480
GTGATTACA CATAATAGGA AACGCTTGGT TTCAGTGAAG TCTGTGTTGT ATATATTCTG CACTAAATGT GTATTATCCT TTGCGAACCA AAGTCACTTC AGACACAAACA TATATAAGAC	3540
TTATATACAC GCATTTGTG TTTGTGTATA TATTCAGT CCATTCAGAT ATGTGTATAT AATATATGTG CGTAAAACAC AAACACATAT ATAAAGTTCA GGTAAGTCTA TACACATATA	3600
AGTGCAGACC TTGTAAATTA AATATTCTGA TACTTTTCC TCAATAAATA TTTAAAT TCACGTCTGG AACATTTAAT TTATAAGACT ATGAAAAAGG AGTTATTTAT AAATTAA	

Figure 6D

SUBSTITUTE SHEET (RULE 26)

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MVCCGPGRML LGWAGLLVLA ALCLLQVPGA QAAACEPVRI PLCKSLPWMM TKMPNHLHHS 60
TQANAILAME QFEGLLGTHC SPDLLFFLCA MYAPICTIDF QHEPIKPKS VCERARQGCE 120
PILIKYRHSH PESLACDELP VYDRGVCISP EAIVTADGAD FPMDSSTGHC RGASSERCKC 180
KPVRATQKTY FRNNNYVIR AKVKEVKMKC HDVTAVVEVK EILKASLVNI PRDTVNLYTT 240
SGCLCPPLTV NEEYVIMGYE DEERSRLLL V EGSI AEKWKD RLGKKVKRWD MKLRHLGLGK 300
TDASDSTQNZ KSGRNSNPRP ARS.

Figure 7
SUBSTITUTE SHEET (RULE 26)

AAGCCTGGGA CCATGGTCTG CTGGGGCCCG GGACGGATGC TGCTAGGATG GGCCGGGTTG	60
TTCGGACCCT GGTACCAGAC GACGCCGGC CCTGCCTACG ACGATCCTAC CGGGCCCAAC	
CTAGTCCTGG CTGCTCTCTG CCTGCTCCAG GTGCCCGGAG CTCAGGCTGC AGCCTGTGAG	120
GATCAGGACC GACGAGAGAC GGACGAGGTC CACGGGCCTC GAGTCCGACG TCGGACACTC	
CCTGTCCGCA TCCCCTGCTG CAAGTCCCTT CCCTGGAACA TGACCAAGAT GCCCAACCAC	180
GGACAGGCGT AGGGCGACAC GTTCAGGGAA GGGACCTTGT ACTGGTTCTA CGGGTTGGTG	
CTGCACCCACA GCACCCAGGC TAACGCCATC CTGGCCATGG AACAGTTCGA AGGGCTGCTG	240
GACGTGGTGT CGTGGGTCCG ATTGCGGTAG GACCGGTACC TTGTCAAGCT TCCCGACGAC	
GGCACCCACT GCAGCCCCGA TCTTCTCTTC TTCCTCTGTG CAATGTACGC ACCCATTG	300
CCGTGGGTGA CGTCGGGCCT AGAAGAGAAG AAGGAGACAC GTTACATGCG TGGTAAACG	
ACCATCGACT TCCAGCACGA GCCCATCAAG CCCTGCAAGT CTGTGTGTGA GCGCGCCCGA	360
TGGTAGCTGA AGGTCGTGCT CGGGTAGTTC GGGACGTTCA GACACACACT CGCGCGGGCT	
CAGGGCTGCG AGCCCATTCT CATCAAGTAC CGCCACTCGT GGCCGGAAAG CTTGGCCTGC	420
GTCCCGACGC TCGGGTAAGA GTAGTTCATG GCGGTGAGCA CCGGCCTTTC GAACCGGACG	
GACGAGCTGC CGGTGTACGA CCGCGGCGTG TGCATCTCTC CTGAGGCCAT CGTCACCGCG	480
CTGCTCGACG GCCACATGCT GGCGCCGCAC ACGTAGAGAG GACTCCGGTA GCAGTGGCGC	
GACGGAGCGG ATTTTCCTAT GGATTCAAGT ACTGGACACT GCAGAGGGGC AAGCAGCGAA	540
CTGCCTCGCC TAAAAGGATA CCTAAGTTCA TGACCTGTGA CGTCTCCCCG TTGTCGCTT	
CGTTGCAAAT GTAAGCCTGT CAGAGCTACA CAGAAGACCT ATTTCCGGAA CAATTACAAC	600
GCAACGTTA CATTGGACA GTCTCGATGT GTCTCTGGA TAAAGGCCTT GTTAATGTG	
TATGTCATCC GGGCTAAAGT TAAAGAGGTA AAGATGAAAT GTCATGATGT GACCGCCGTT	660
ATACAGTAGG CCCGATTCA ATTTCTCCAT TTCTACTTTA CAGTACTACA CTGGCGGCAA	
GTGGAAGTGA AGGAAATTCT AAAGGCATCA CTGGTAAACA TTCCAAGGGGA CACCGTCAAT	720
CACCTTCACT TCCTTTAAGA TTTCCGTAGT GACCATTGT AAGGTTCCCT GTGGCAGTTA	
CTTTATACCA CCTCTGGCTG CCTCTGTCCCT CCACTTACTG TCAATGAGGA ATATGTCATC	780
GAAATATGGT GGAGACCGAC GGAGACAGGA GGTGAATGAC AGTTACTCCT TATACAGTAG	
ATGGGCTATG AAGACGAGGA ACGTTCCAGG TTACTCTTGG TAGAAGGCTC TATAGCTGAG	840
TACCCGATAC TTCTGCTCCT TGCAAGGTCC AATGAGAACC ATCTTCCGAG ATATCGACTC	
AAGTGGAAAGG ATCGGCTTGG TAAGAAAGTC AAGCGCTGGG ATATGAAACT CCGACACCTT	900
TTCACCTTCC TAGCCGAACC ATTCTTCAG TTGCGGACCC TATACTTTGA GGCTGTGGAA	
GGACTGGGTA AAACTGATGC TAGCGATTCC ACTCAGAACG AGAAGTCTGG CAGGAACCT	960
CCTGACCCAT TTTGACTACG ATCGCTAAGG TGAGTCTTAG TCTTCAGACC GTCCCTTGAGA	

AATCCCCGGC CAGCACCGCAG CTAAATCCTG AAATGTAAAA GGCCACACCC ACGGACTCCC	1020
TTAGGGGCCG GTCGTGCGTC GATTTAGGAC TTTACATTTT CCGGTGTGGG TGCCCTGAGGG	
TTCTAAGACT GGCGCTGGTG GACTAACAAA GGAAAACCGC ACAGTTGTGC TCGTGACCGA	1080
AAGATTCTGA CCGCGACCAC CTGATTGTTT CCTTTGGCG TGTCAACACG AGCACTGGCT	
TTGTTTACCG CAGACACCGC GTGGCTACCG AAGTTACTTC CGGTCCCCTT TCTCCTGCTT	1140
AACAAATGGC GTCTGTGGCG CACCGATGGC TTCAATGAAG GCCAGGGAA AGAGGACGAA	
CTTAATGGCG TGGGGTTAGA TCCTTTAATA TGTTATATAT TCTGTTCAT CAATCACGTG	1200
GAATTACCGC ACCCCAAATCT AGGAAATTAT ACAATATATA AGACAAAGTA GTTAGTGCAC	
GGGACTGTTC TTTTGCAACC AGAATAGTAA ATTAAATATG TTGATGCTAA GGTTTCTGTA	1260
CCCTGACAAG AAAACGTTGG TCTTATCATT TAATTATAC AACTACGATT CCAAAGACAT	
CTGGACTCCC TGGGTTTAAT TTGGTGTCT GTACCCCTGAT TGAGAATGCA ATGTTTCATG	1320
GACCTGAGGG ACCCAAATTA AACCAACAAGA CATGGGACTA ACTCTTACGT TACAAAGTAC	
TAAAGAGAGA ATCCTGGTCA TATCTCAAGA ACTAGATATT GCTGTAAGAC AGCCTCTGCT	1380
ATTTCTCTCT TAGGACCACT ATAGAGTTCT TGATCTATAA CGACATTCTG TCGGAGACGA	
GCTGCGCTTA TAGTCTTGTG TTTGTATGCC TTTGTCCATT TCCCTCATGC TGTGAAAGTT	1440
CGACGCGAAT ATCAGAACAC AAACATACGG AAACAGGTAA AGGGAGTACG ACACTTCAA	
ATACATTTT ATAAAGGTAG AACGGCATT TGAAATCAGA CACTGCACAA GCAGAGTAGC	1500
TATGTACAAA TATTTCCATC TTGCCGTAAA ACTTTAGTCT GTGACGTGTT CGTCTCATCG	
CCAACACCAAG GAAGCATTG TGAGGAAACG CCACACAGCA TGACTTATT TCAAGATTGG	1560
GGTTGTGGTC CTTCGTAAAT ACTCCTTTGC GGTGTGTCGT ACTGAATAAA AGTTCTAAC	
CAGGCAGCAA AATAAATAGT GTTGGGAGCC AAGAAAAGAA TATTTTGCTT GGTTAAGGGG	1620
GTCCGTGTT TTATTTATCA CAACCCTCGG TTCTTTCTT ATAAAACGGA CCAATTCCCC	
CACACTGGAA TCAGTAGCCC TTGAGCCATT AACAGCAGTG TTCTTCTGGC AAGTTTTGA	1680
GTGTGACCTT AGTCATCGGG AACTCGGTAA TTGTCGTAC AAGAAGACCG TTCAAAAACT	
TTTGTTCATA AATGTATTCA CGAGCATTAG AGATGAACCT ATAACTAGAC ATCTGTTGTT	1740
AAACAAGTAT TTACATAAGT GCTCGTAATC TCTACTTGAA TATTGATCTG TAGACAACAA	
ATCTCTATAG CTCTGCTTCC TTCTAAATCA AACCCATTGT TGGATGCTCC CTCTCCATTG	1800
TAGAGATATC GAGACGAAGG AAGATTTAGT TTGGGTAACA ACCTACGAGG GAGAGGTAAG	

Figure 8B
SUBSTITUTE SHEET (RULE 26)

ATAAATAAAT TTGGCTTGCT GTATTGGCCA GGAAAAGAAA GTATTAAAGT ATGCATGCAT 1860
TATTATTTA AACCGAACGA CATAACCGGT CCTTTCTTT CATAATTCA TACGTACGTA

GTGCACCAGG GTGTTATTTA ACAGAGGTAT GTAACTCTAT AAAAGACTAT AATTTACAGG 1920
CACGTGGTCC CACAATAAAT TGTCTCCATA CATTGAGATA TTTTCTGATA TTAAATGTCC

ACACGGAAAT GTGCACATTT GTTACTTTT TTTCTTCCTT TTGCTTTGGG CTTGTGATTT 1980
TGTGCCTTTA CACGTGTAAA CAAATGAAAA AAAGAAGGAA AACGAAACCC GAACACTAAA

TGGTTTTGG TGTGTTATG TCTGTATTTT GGGGGGTGGG TAGGTTAAG CCATTGCACA 2040
ACCAAAAAACC ACACAAATAC AGACATAAAA CCCCCCACCC ATCCAAATTC GGTAACGTGT

TTCAAGTTGA ACTAGATTAG AGTAGACTAG GCTCATTGGC CTAGACATTA TGATTTGAAT 2100
AAGTTCAACT TGATCTAATC TCATCTGATC CGAGTAACCG GATCTGTAAT ACTAAACTTA

TTGTGTTGTT TAATGCTCCA TCAAGATGTC TAATAAAAGG AATATGGTTG TCAACAGAGA 2160
AACACAAACAA ATTACGAGGT AGTTCTACAG ATTATTTCC TTATACCAAC AGTTGTCTCT

CGACAAACAAC AACAAA
GCTGTTGTTG TTGTTT

MVCGSPGGML LLRAGLLALA ALCLLRVPGA RAAACEPVRI PLCKSLPWNM TKMPNHLHHS	60
TQANAILAIE QFEGLLGTHC SPDLLFFLCA MYAPICTIDF QHEPIKPKS VCERARQGCE	120
PILIKYRHSW PENLACEELP VYDRGVCISP EAIVTADGAD FPMDSNGNC RGASSERCKC	180
KPIRATQKTY FRNNNYVIR AKVKEIKTKC HDVTAVVEVK EILKSSLVNI PRDTVNLYTS	240
SGCLCPPLNV NEEYIIMGYE DEERSRLLL V EGSI AEKWKD RL GKKV KRWD MKL RHLGLSK	300
SDSSNSDSTQ SQKSGRNSNP RQARN.	

GGCGGAGCGG GCCTTTGGC GTCCACTGCG CGGCTGCACC CTGCCCATC TGCCGGATC CCGCCTCGCC CGGAAAACCG CAGGTGACGC GCCGACGTGG GACGGGGTAG ACGGCCCTAG	60
ATGGTCTGCG GCAGCCCGGG AGGGATGCTG CTGCTGCAGG CCGGGCTGCT TGCCCTGGCT TACCAGACGC CGTCGGGCCCG TCCCTACGAC GACGACGCCG GGCCGACGA ACAGGGACCGA	120
GCTCTCTGCC TGCTCCGGGT GCCCGGGCT CGGGCTGCAG CCTGTGAGCC CGTCCGCATC CGAGAGACGG ACGAGGCCCA CGGGCCCCGA GCCGACGTC GGACACTCGG GCAGGCGTAG	180
CCCCTGTGCA AGTCCCTGCC CTGGAACATG ACTAAGATGC CCAACCACCT GCACCACAGC GGGGACACGT TCAGGGACGG GACCTTGTAC TGATTCTACG GGTTGGTGGA CGTGGTGTG	240
ACTCAGGCCA ACGCCATCCT GGCCATCGAG CAGTTCGAAG GTCTGCTGGG CACCCACTGC TGAGTCCGGT TGCGGTAGGA CCGGTAGCTC GTCAAGCTTC CAGACGACCC GTGGGTGACG	300
AGCCCCGATC TGCTCTTCTT CCTCTGTGCC ATGTACGCC CCATCTGCAC CATTGACTTC TCGGGGCTAG ACGAGAAGAA GGAGACACGG TACATGCGCG GGTAGACGTG GTAAGTGAAG	360
CAGCACCGAGC CCATCAAGCC CTGTAAGTCT GTGTGCGAGC GGGCCCGGCA GGGCTGTGAG GTCGTGCTCG GGTAGTTCGG GACATTAGA CACACGCTCG CCCGGCCGT CCCGACACTC	420
CCCATACTCA TCAAGTACCG CCACTCGTGG CCGGAGAACCC TGGCCTGCGA GGAGCTGCCA GGGTATGAGT AGTTCATGGC GGTGAGCACC GGCTCTTGG ACCGGACGCT CCTCGACGGT	480
GTGTACGACA GGGCGTGTG CATCTCTCCC GAGGCCATCG TTACTGCGGA CGGAGCTGAT CACATGCTGT CCCCACACAC GTAGAGAGGG CTCCGGTAGC AATGACGCC GCCTCGACTA	540
TTTCCTATGG ATTCTAGTAA CGGAAACTGT AGAGGGCAA GCAGTGAACG CTGTAAATGT AAAGGATACC TAAGATCATT GCCTTTGACA TCTCCCCGTT CGTCACTTGC GACATTACAA	600
AAGCCTATTA GAGCTACACA GAAGACCTAT TTCCGGAACA ATTACAACCA TGTCATTGG TTCCGATAAT CTCGATGTGT CTTCTGGATA AAGGCCTTGT TAATGTTGAT ACAGTAAGCC	660
GCTAAAGTTA AAGAGATAAA GACTAAGTGC CATGATGTGA CTGCAGTAGT GGAGGTGAAG CGATTCAAT TTCTCTATTT CTGATTCAAG GTACTACACT GACGTCACTA CCTCCACTTC	720
GAGATTCTAA AGTCCTCTCT GTAAACATT CCACGGGACA CTGTCAACCT CTATACCAGC CTCTAAGATT TCAGGAGAGA CCATTGTAA GGTGCCCTGT GACAGTTGGA GATATGGTCG	780
TCTGGCTGCC TCTGCCCTCC ACTTAATGTT AATGAGGAAT ATATCATCAT GGGCTATGAA AGACCGACGG AGACGGGAGG TGAATTACAA TTACTCCTTA TATAGTAGTA CCCGATACTT	840

Figure 10A
SUBSTITUTE SHEET (RULE 26)

GATGAGGAAC GTTCCAGATT ACTCTGGTG GAAGGCTCTA TAGCTGAGAA GTGGAAGGAT 900
 CTACTCCTTG CAAGGTCTAA TGAGAACAC CTTCCGAGAT ATCGACTCTT CACCTTCCTA

 CGACTCGGTA AAAAAGTTAA GCGCTGGGAT ATGAAGCTTC GTCATCTTGG ACTCAGTAAA 960
 GCTGAGCCAT TTTTCATT CGCGACCCTA TACTTCGAAG CAGTAGAACC TGAGTCATTT

AGTGATTCTA GCAATAGTGA TTCCACTCAG AGTCAGAAGT CTGGCAGGAA CTCGAACCCC 1020
TCACTAAGAT CGTTATCACT AAGGTGAGTC TCAGTCTTC GACCGTCCTT GAGCTTGGGG

 CGGCAAGCAC GCAACTAAAT CCCGAAATAC AAAAAGTAAC ACAGTGGACT TCCTATTAAG 1080
 GCCGTTCGTG CGTTGATTAA GGGCTTATG TTTTCATTG TGTACCTGA AGGATAATTC

 ACTTACTTGC ATTGCTGGAC TAGCAAAGGA AAATTGCACT ATTGCACATC ATATTCTATT 1140
 TGAATGAACG TAACGACCTG ATCGTTTCCT TTTAACGTGA TAACGTGTAG TATAAGATAAA

 GTTTACTATA AAAATCATGT GATAACTGAT TATTACTTCT GTTTCTCTTT TGGTTTCTGC 1200
 CAAATGATAT TTTAGTACA CTATTGACTA ATAATGAAGA CAAAGAGAAA ACCAAAGACG

 TTCTCTCTTC TCTCAACCCC TTTGTAATGG TTTGGGGCA GACTCTTAAG TATATTGTGA 1260
 AAGAGAGAAG AGAGTTGGGG AAACATTACC AAACCCCCGT CTGAGAATTTC ATATAACACT

 GTTTTCTATT TCACTAATCA TGAGAAAAAC TGTTCTTTG CAATAATAAT AAATTAAACA 1320
 CAAAGATAA AGTGATTAGT ACTCTTTTG ACAAGAAAAC GTTATTATTA TTTAATTGT

 TGCTGTTACC AGAGCCTCTT TGCTGAGTCT CCAGATGTTA ATTACTTTC TGCACCCCAA 1380
 ACGACAATGG TCTCGGAGAA ACGACTCAGA GGTCTACAAT TAAATGAAAG ACGTGGGGTT

 TTGGGAATGC AATATTGGAT GAAAAGAGAG GTTTCTGGTA TTCACAGAAA GCTAGATATG 1440
 AACCCCTTACG TTATAACCTA CTTTCTCTC CAAAGACCCT AAGTGTCTT CGATCTATAC

 CCTTAAACA TACTCTGCCG ATCTAATTAC AGCCTTATT TTGTATGCCT TTTGGCATT 1500
 GGAATTGT ATGAGACGGC TAGATTAATG TCGGAATAAA AACATACGGA AAACCCGTAA

 CTCCTCATGC TTAGAAAGTT CCAAATGTTT ATAAAGGTAA AATGGCAGTT TGAAGTCAA 1560
 GAGGAGTACG AATCTTCAA GGTTACAAA TATTCCATT TTACCGTCAA ACTTCAGTTT

 TGTACATAG GCAAAGCAAT CAAGCACCAAG GAAGTGTAA TGAGGAAACA ACACCCAAGA 1620
 ACAGTGTATC CGTTCTGTTA GTTCGTGGTC CTTCACAAAT ACTCCTTGT TGTGGGTTCT

 TGAATTATT TTGAGACTGT CAGGAAGTAA AATAAATAGG AGCTTAAGAA AGAACATT 1680
 ACTTAATAAA AACTCTGACA GTCCTTCATT TTATTATCC TCGAATTCTT TCTTGTAAAA

 GCCTGATTGA GAAGCACAAAC TGAAACCAAGT AGCCGCTGGG GTGTTAATGG TAGCATTCTT 1740
 CGGACTAACT CTTCGTGTG ACTTTGGTCA TCGGCGACCC CACAATTACC ATCGTAAGAA

 CTTTGGCAA TACATTGAT TTGTTCATGA ATATATTAAT CAGCATTAGA GAAATGAATT 1800
 GAAAACCGTT ATGTAAACTA AACAAAGTACT TATATAATTAA GTCGTAATCT CTTTACTTAA

 ATAACCTAGAC ATCTGCTGTT ATCACCAAG TTTGTTAA TTTGCTTCCT TTAAATAAA 1860
 TATTGATCTG TAGACGACAA TAGTGGTATC AAAACAAATT AACAGAAGGA AAATTATTT

 CCCATTGGTG AAAGTCAAAA AAAAAAAA AAA
 GGGTAACCAC TTTCAGTTT TTTTTTTTT TTT

Figure 10B
 SUBSTITUTE SHEET (RULE 26)

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US97/10942

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) :Please See Extra Sheet.

US CL : 530/300, 350; 514/2; 536/23.1

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 530/300, 350; 514/2; 536/23.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

DIALOG (MEDLINE, BIOSIS, EMBASE, WPI, USPATFULL) AUTHOR AND WORD. search terms: e.g. cerberus, xenopus

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y, P	BOUWMEESTER et al. Cerberus is a head-inducing secreted factor expressed in the anterior endoderm of Spemann's organizer. Nature. 15 August 1996, Vol. 382, No. 6592, pages 595-601, see entire document.	1-15

Further documents are listed in the continuation of Box C. See patent family annex.

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Date of the actual completion of the international search

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Date of mailing of the international search report

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A. CLASSIFICATION OF SUBJECT MATTER:

IPC (6):

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